



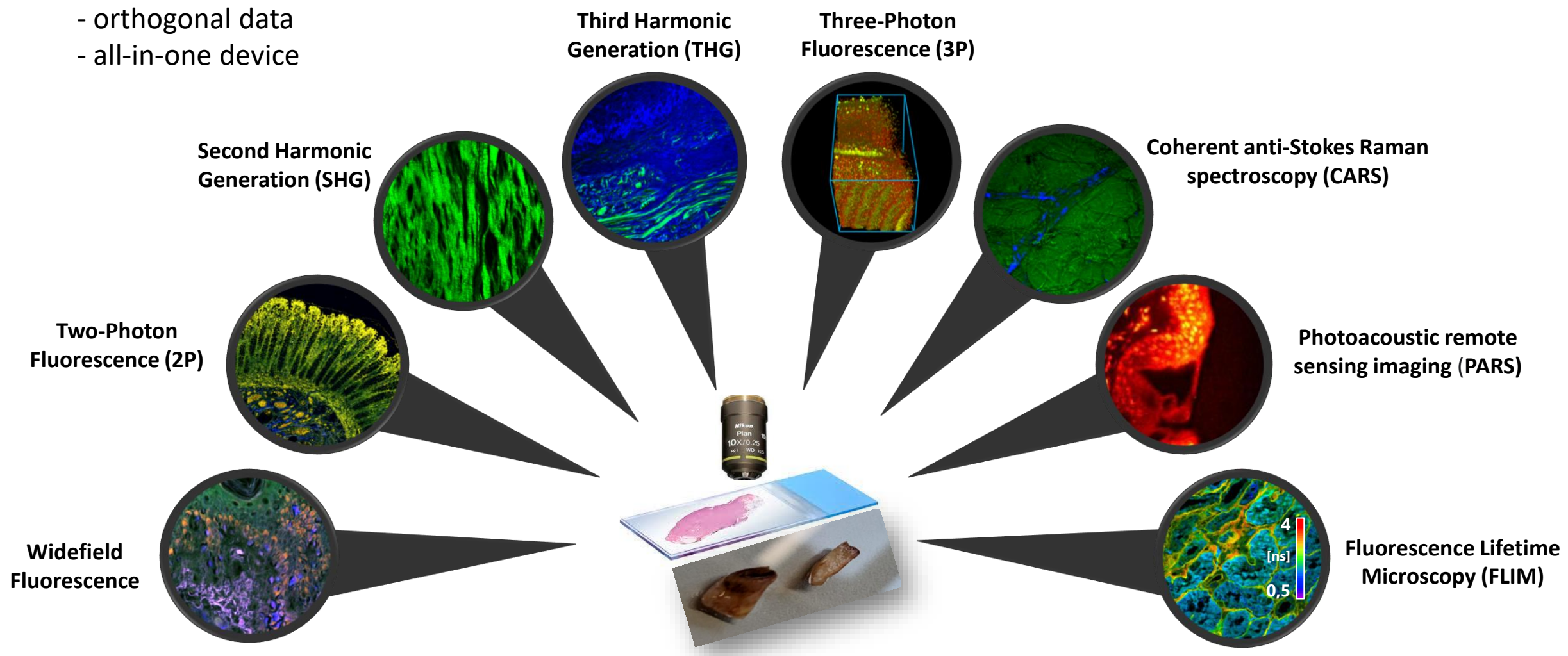
**PROSPECTIVE  
INSTRUMENTS**

MULTIPHOTON MICROSCOPES  
FEMTOSECOND LASERS  
CONTRACT IMAGING SERVICE

[WWW.P-INST.COM](http://WWW.P-INST.COM)

# MULTIMODAL IMAGING VISION

- non destructive
- orthogonal data
- all-in-one device



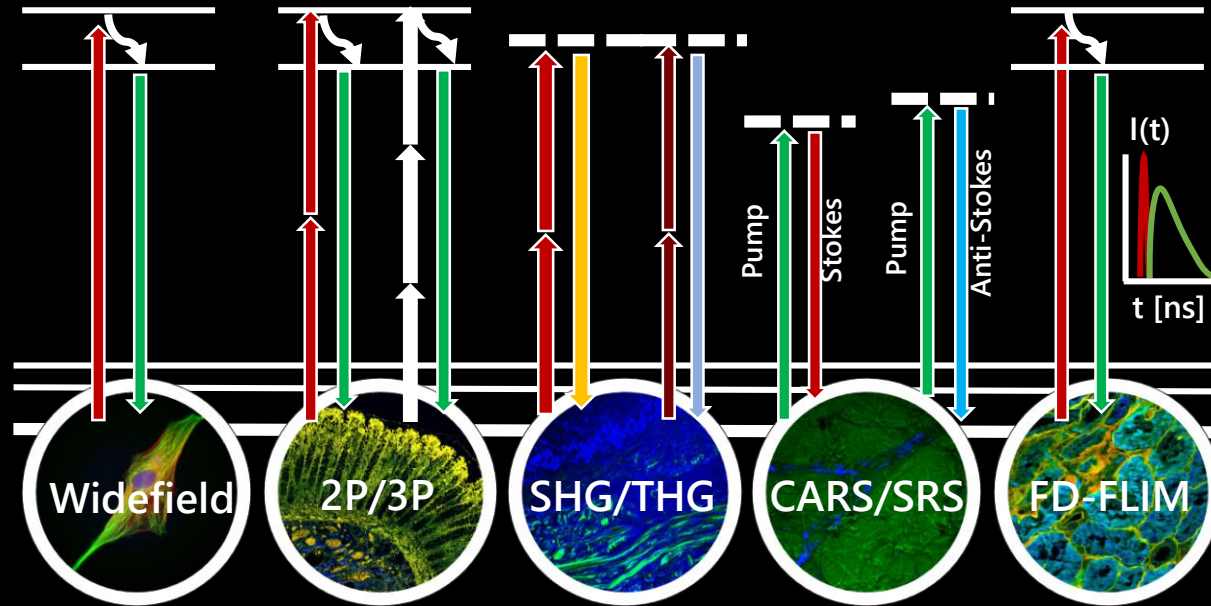


PROSPECTIVE  
LIFE SCIENCE



TURN-KEY  
EASY-TO-USE  
COMPACT  
MULTIMODAL

# MULTIMODAL MICROSCOPY



PROSPECTIVE  
LASERS



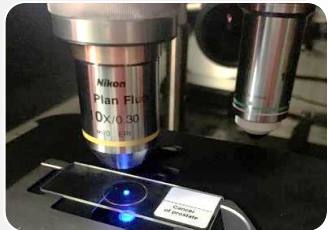
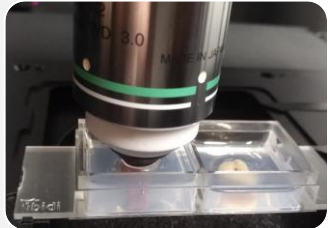
TWO-PHOTON, SHG & THG  
OPTOGENETICS  
FLIM  
CARS & SRS

Multiphoton Microscopes

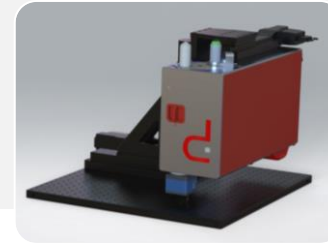
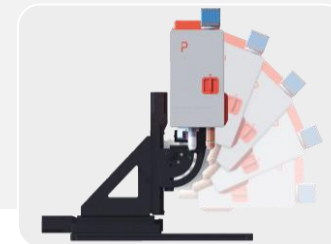
Femtosecond Lasers

# MULTIPHOTON MICROSCOPES

MP-SERIES



- Turn-key, Easy-to-use & rugged
- Compact & air-cooled
- Built in Laser (MPX) or 3P-ready (MPC)
- Multimodal (WF, 2P, SHG, CARS, FLIM)
- Large working distance, 360° access
- Upright & inverted
- Oblique angles

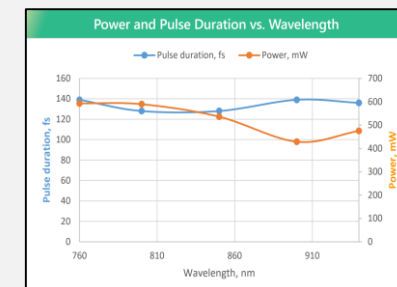


# FEMTOSECOND LASERS

FSX-SERIES



- Turn-key & rugged
- Compact & air-cooled
- Dual-output
- Wavelength tuneable
- High power
- Low RIN & phase noise



- 2P polymerization
- Supercontinuum generation
- Terahertz generation
- Quantum optics
- Amplifier seeding
- 2P/SHG/THG microscopy
- FLIM microscopy
- CARS/SRS microscopy
- Spectroscopy
- Neuroscience & Optogenetics

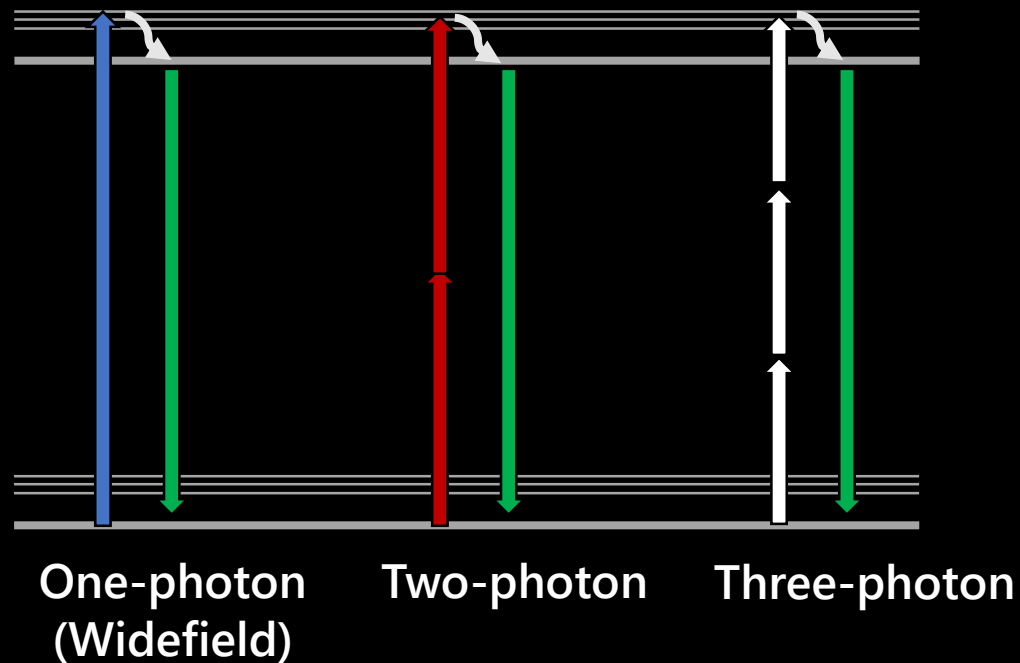
# 2PEF/3PEF VS. WIDEFIELD



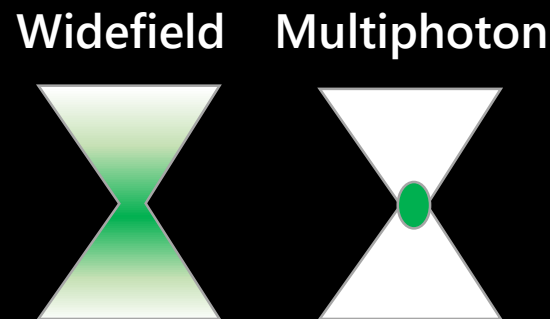
1. 2P/3P excitation is intrinsic confocal due to a confined excitation volume in 3D space
2. 2P/3P excitation is sample saving: only fluorescence in focal plane and point scanning
3. 2PEF/3PEF uses NIR light for higher penetration depth

# MULTIPHOTON EXCITATION FLUORESCENCE

## EXCITATION & ABSORPTION



For non-linear microscopy techniques:  $\text{signal} \propto \text{Power}^n$   
→ multiphoton absorption is spatially confined to the perifocal region. The absence of multiphoton absorption in out-of-focus planes makes this technique **intrinsic confocal!**

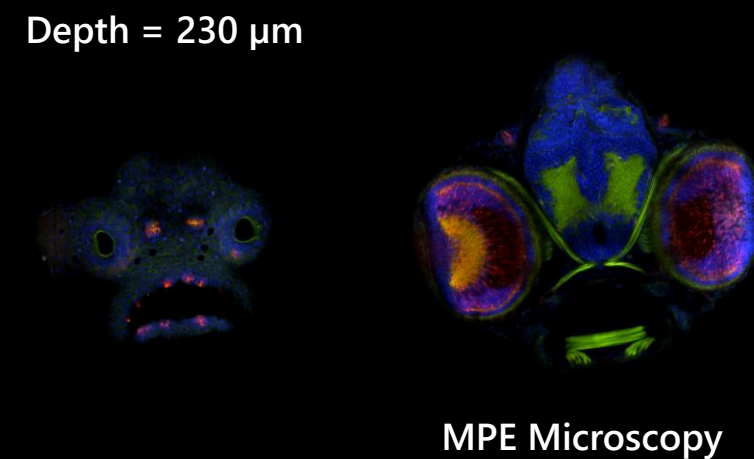
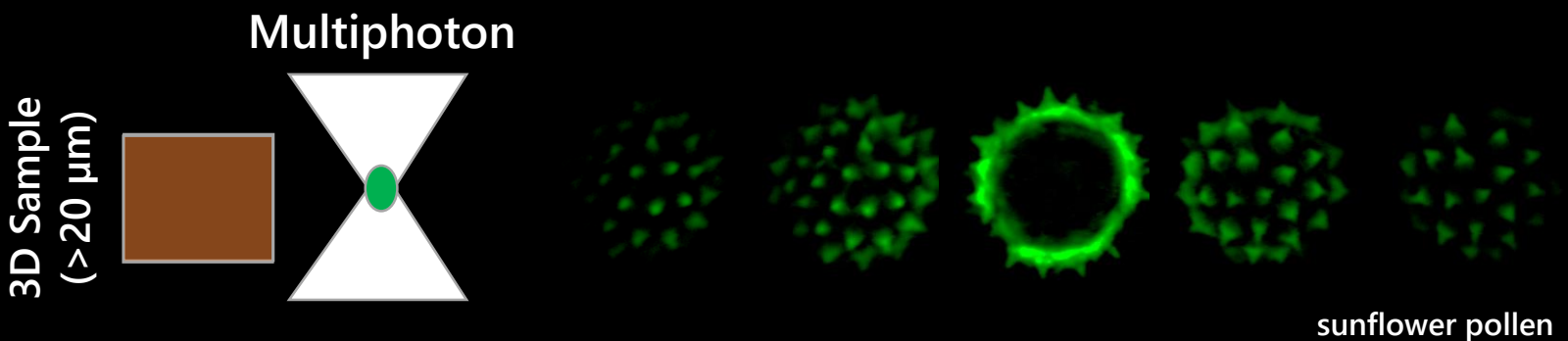
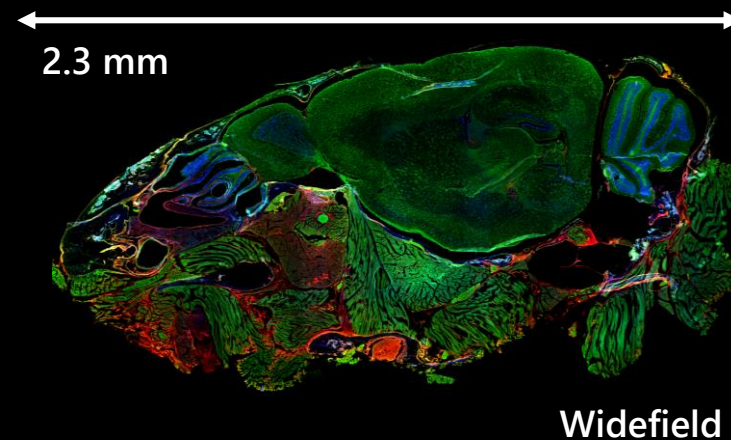
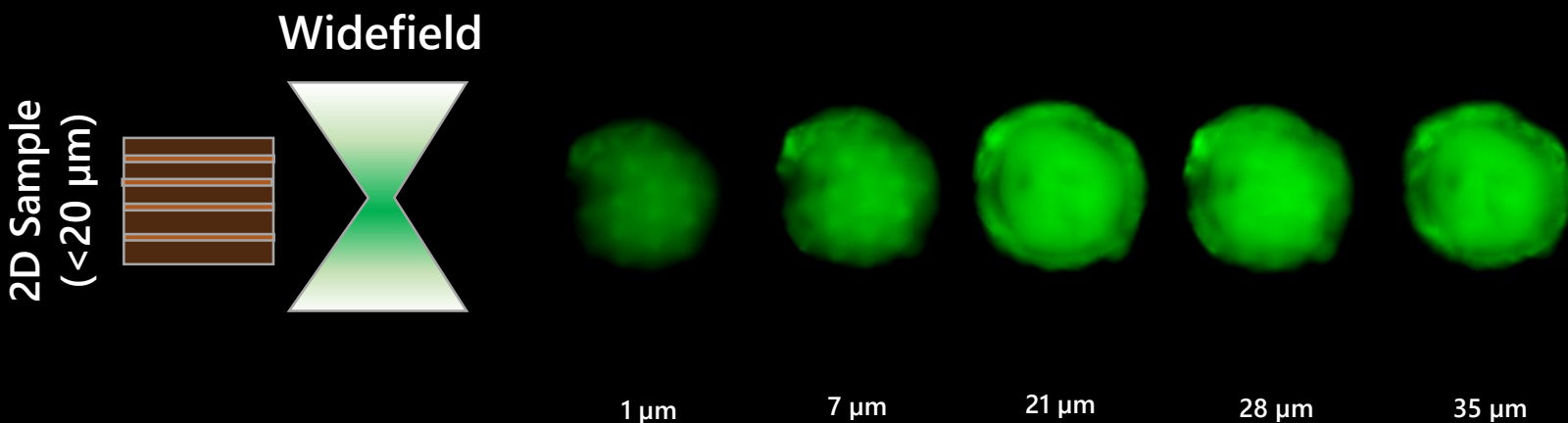


\*Drobizhev, M. et al. Two-photon absorption properties of fluorescent proteins. *Nat Methods* 8, 393–399 (2011)

\*\*Helmchen F, Denk W. Deep tissue two-photon microscopy. *Nat Methods*. 2005 Dec;2(12)

# MULTIPHOTON EXCITATION FLUORESCENCE

OPTICAL SECTIONING

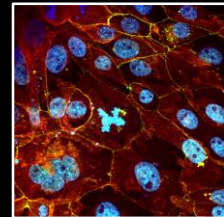
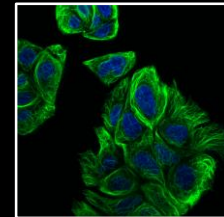
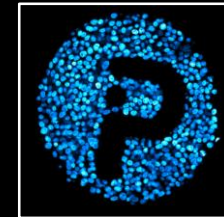
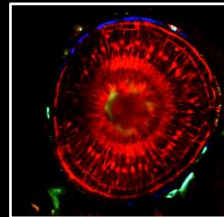
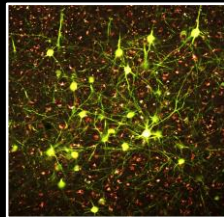
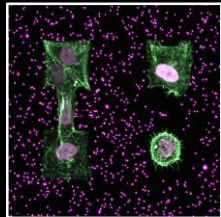
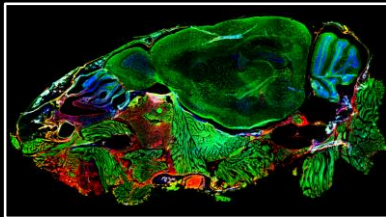
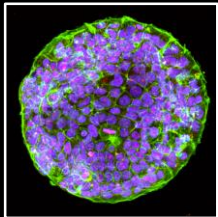
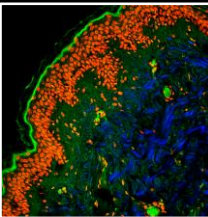
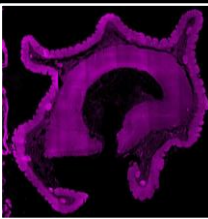
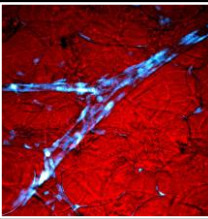
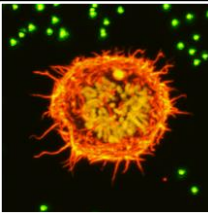




# BROAD RANGE OF APPLICATIONS



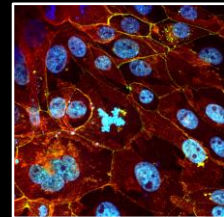
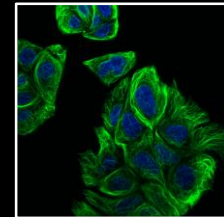
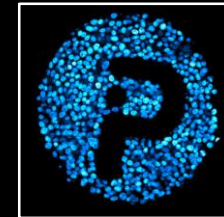
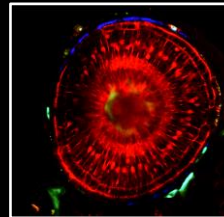
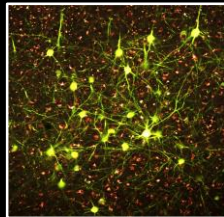
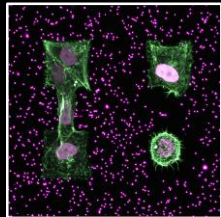
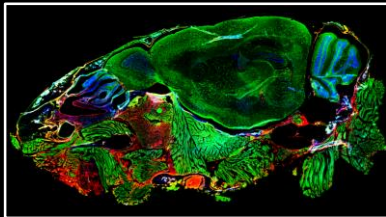
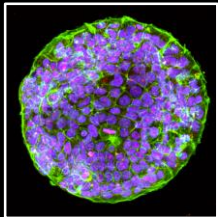
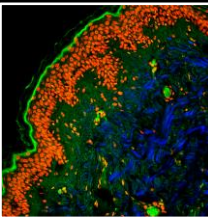
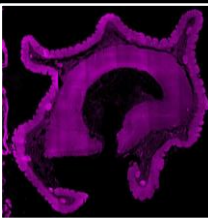
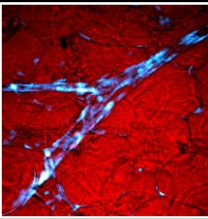
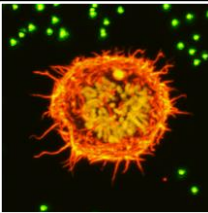
- 2D/3D/4D imaging
- In-vitro & in-vivo
- Whole slide imaging
- Label-free & IHC/ICC
- Non-destructive
- Deep tissue imaging
- Pathology & Cancer
- Neuroscience
- Optogenetics
- Tissue Engineering & Bioprinting
- Spheroids / Organoids



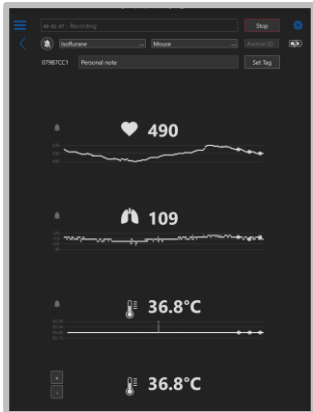
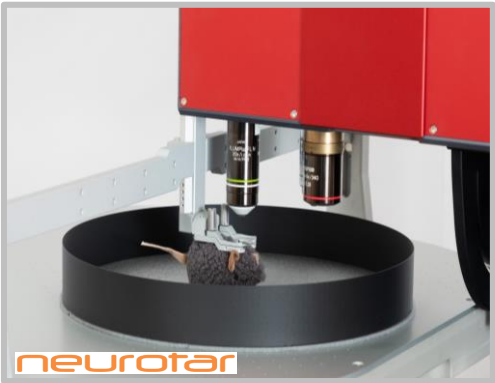
# BROAD RANGE OF APPLICATIONS



- 2D/3D/4D imaging
- **In-vitro & in-vivo**
- Whole slide imaging
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- **Deep tissue imaging**
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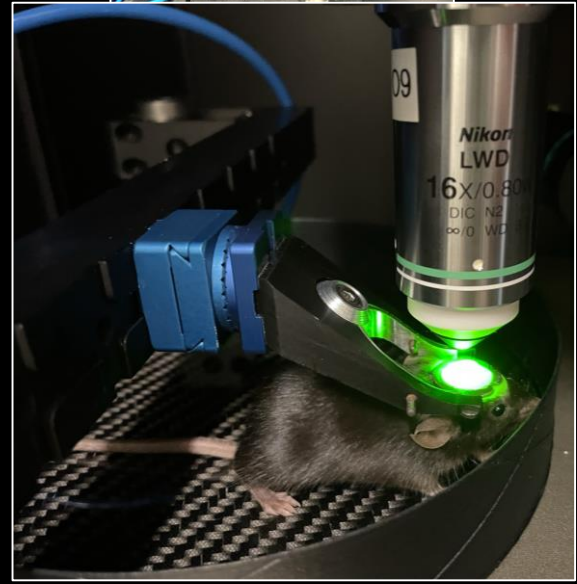
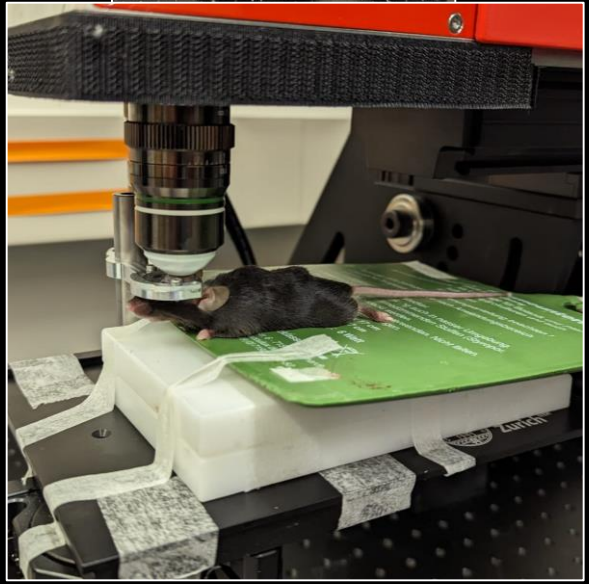
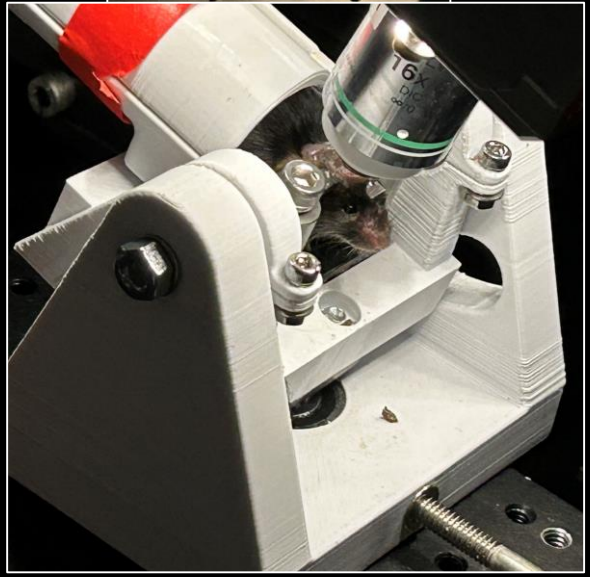
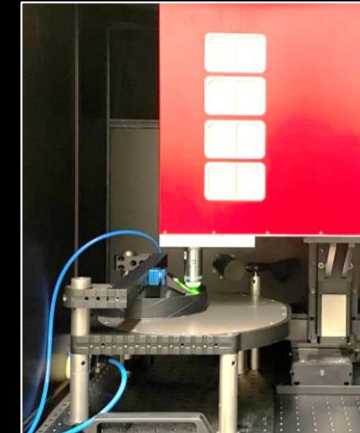
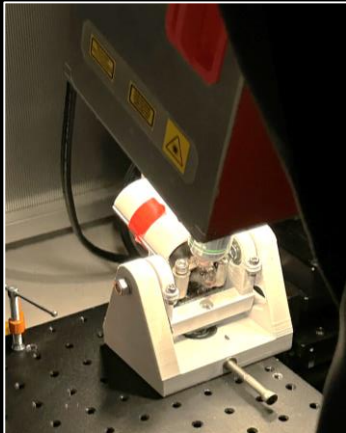


# MPX NEUROEXPLORER



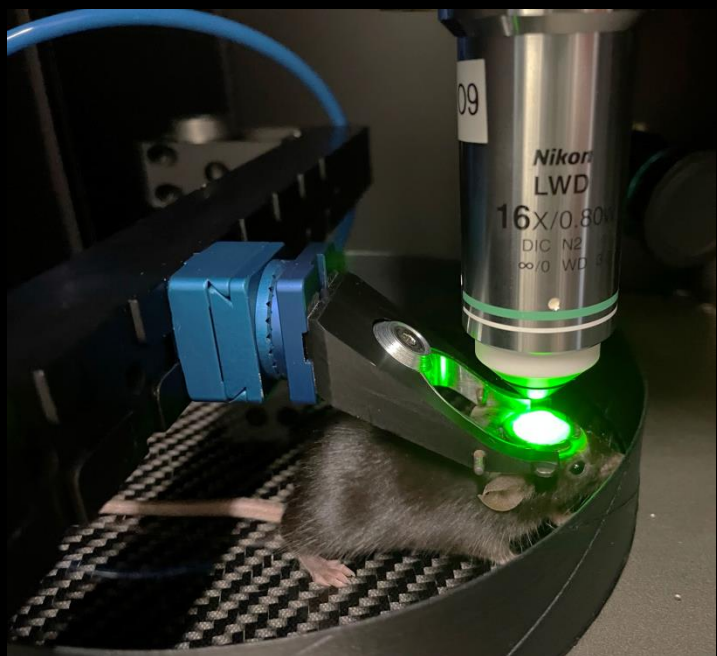


# MPX NEUROEXPLORER

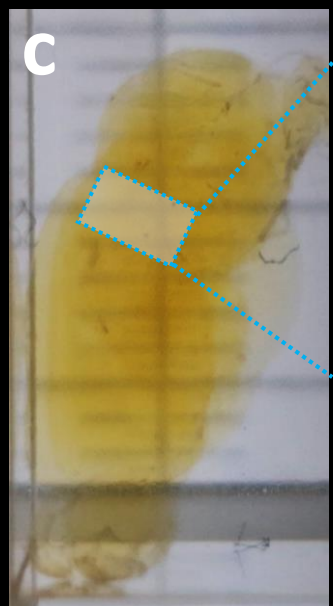
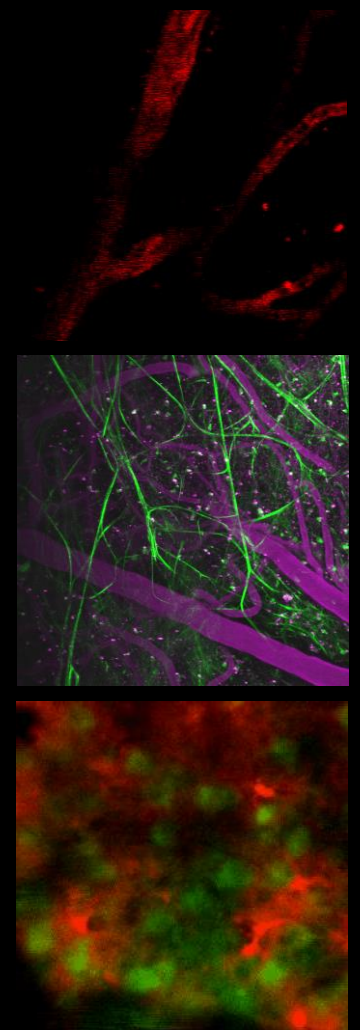




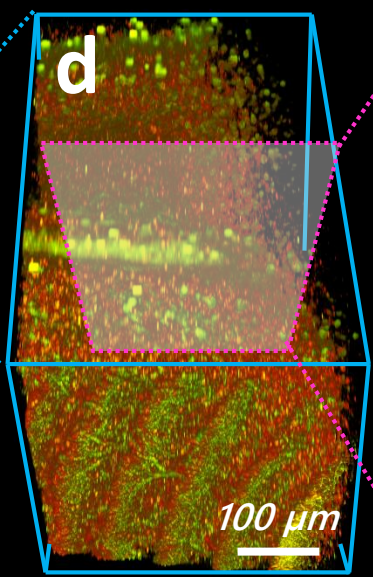
# MPX NEUROEXPLORER: IN-VIVO & WHOLE BRAIN IMAGING



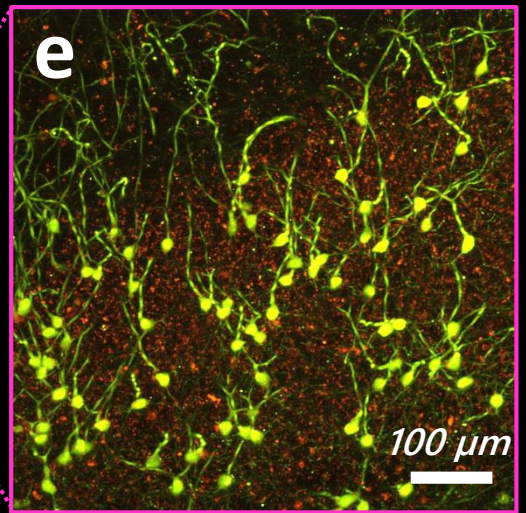
Microscope setup of live mouse in-vivo brain imaging



Whole cleared mouse brain



Volume scan of the cleared mouse brain with a total depth of ~3.5 mm in depth

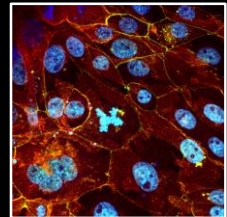
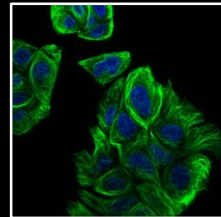
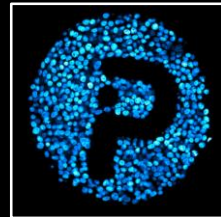
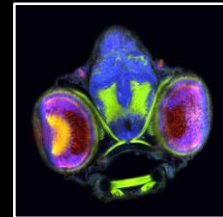
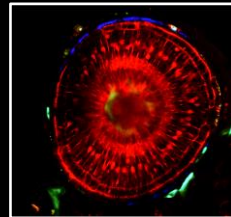
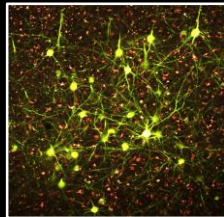
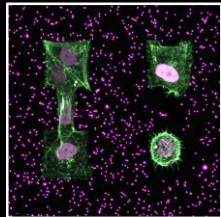
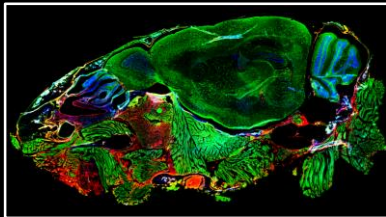
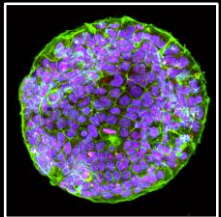
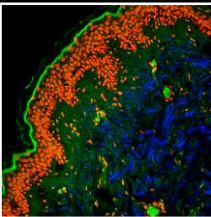
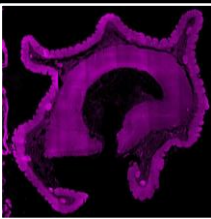
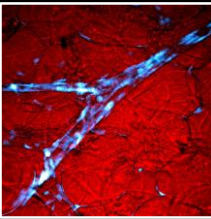
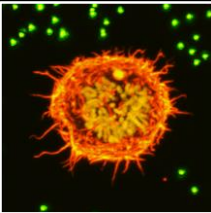


Exemplary z-plane showing stained neurons

# BROAD RANGE OF APPLICATIONS



- 2D/3D/4D imaging
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- **Whole slide imaging**
- **Label-free & IHC/ICC**
- **Non-destructive**
- Deep tissue imaging
- **Pathology & Cancer**
- Neuroscience
- Optogenetics
- Tissue Engineering & Bioprinting
- Spheroids / Organoids



## EXAMPLE: LABEL-FREE BIOMARKERS

**BIOMARKER:** „measurable indicator of some biological state or condition“ (Wikipedia)

<b>Optical properties:</b>	e.g. fluorescence, absorption
<b>Molecular properties:</b>	e.g. metabolic state
<b>Mechanical properties:</b>	e.g. stiffness
<b>Morphological properties:</b>	e.g. cell nucleus
<b>Structural properties:</b>	e.g. wall thickness

**Biomarkers in medicine:**

*Predictive, diagnostic, prognostic indicator*

**Biomarkers in Research:**

*“Contact-free, non-destructive”*

## EXAMPLE: LABEL-FREE BIOMARKERS

**BIOMARKER:** „measurable indicator of some biological state or condition“ (Wikipedia)

<b>Optical properties:</b>	e.g. fluorescence, absorption
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<b>Structural properties:</b>	e.g. wall thickness

**Biomarkers in medicine:**

*Predictive, diagnostic, prognostic indicator*

**Biomarkers in Research:**

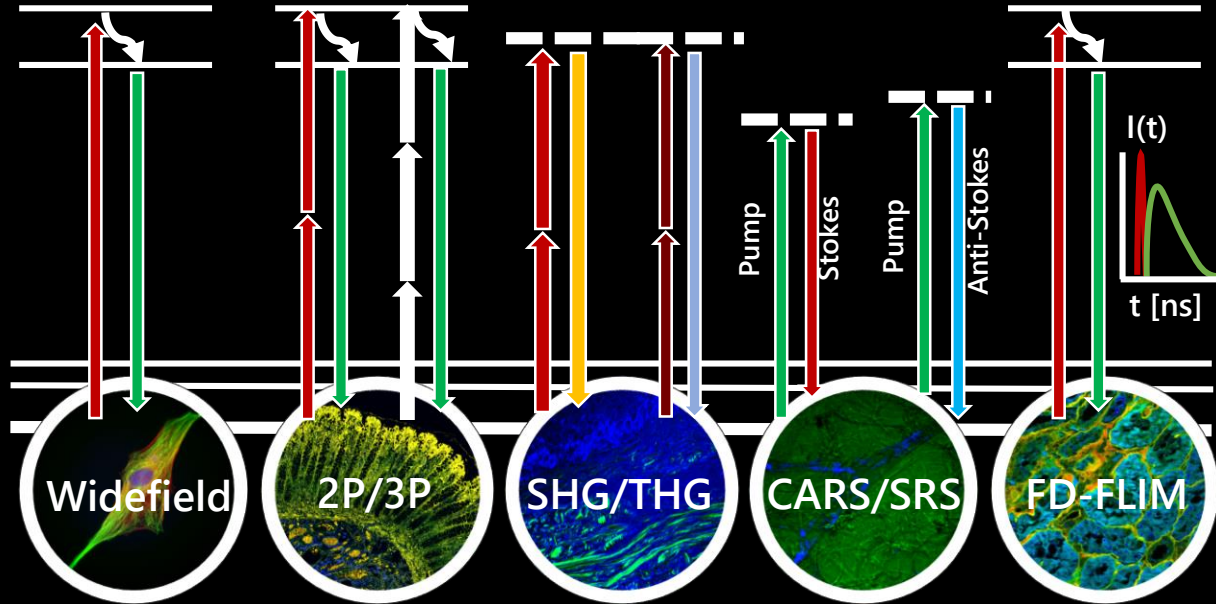
*“Contact-free, non-destructive”*



# MULTIMODAL MICROSCOPY



**TURN-KEY  
EASY-TO-USE  
COMPACT  
MULTIMODAL**

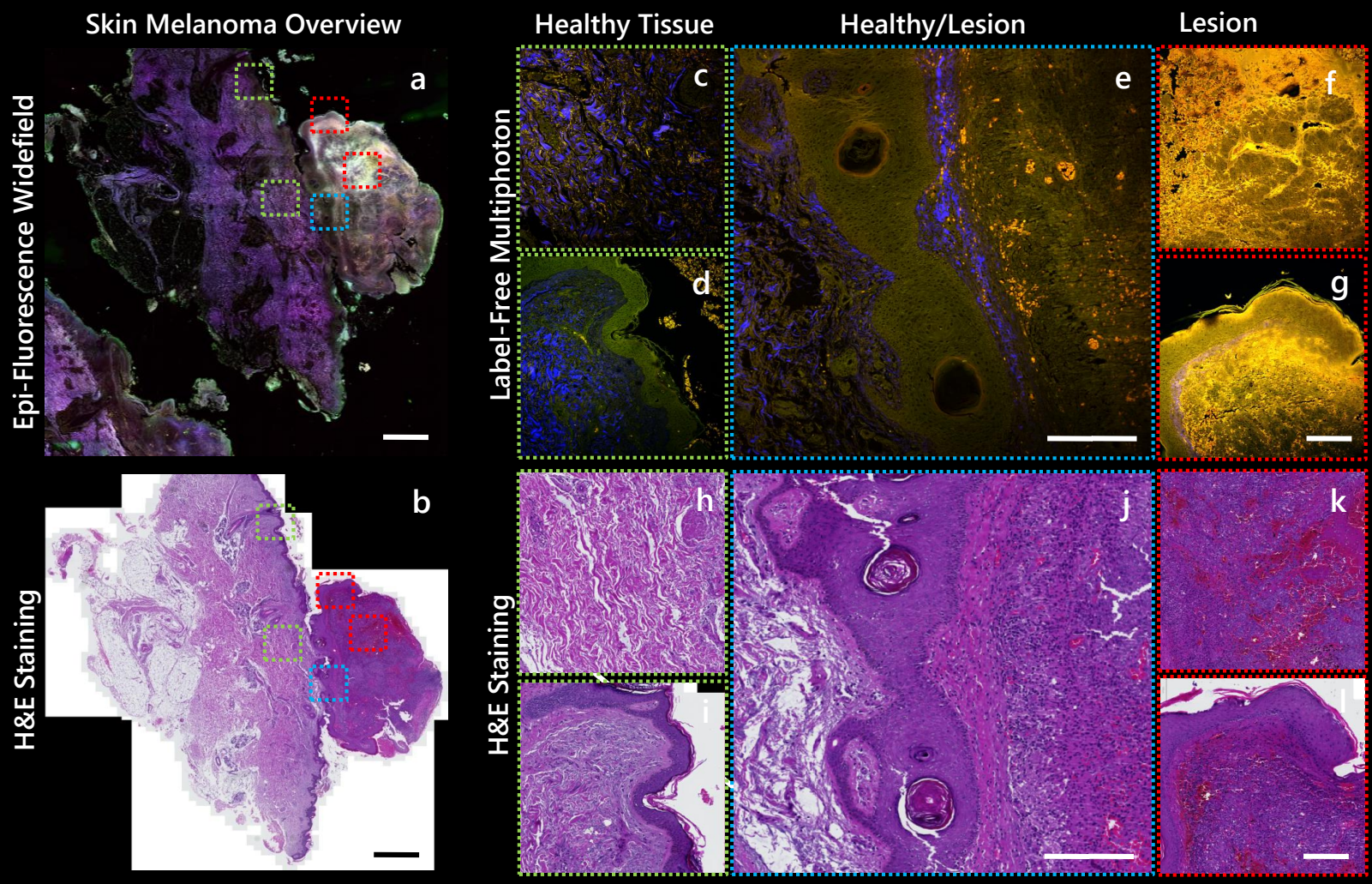


**TWO-PHOTON, SHG & THG  
OPTOGENETICS  
FLIM  
CARS & SRS**

Multiphoton Microscopes

Femtosecond Lasers

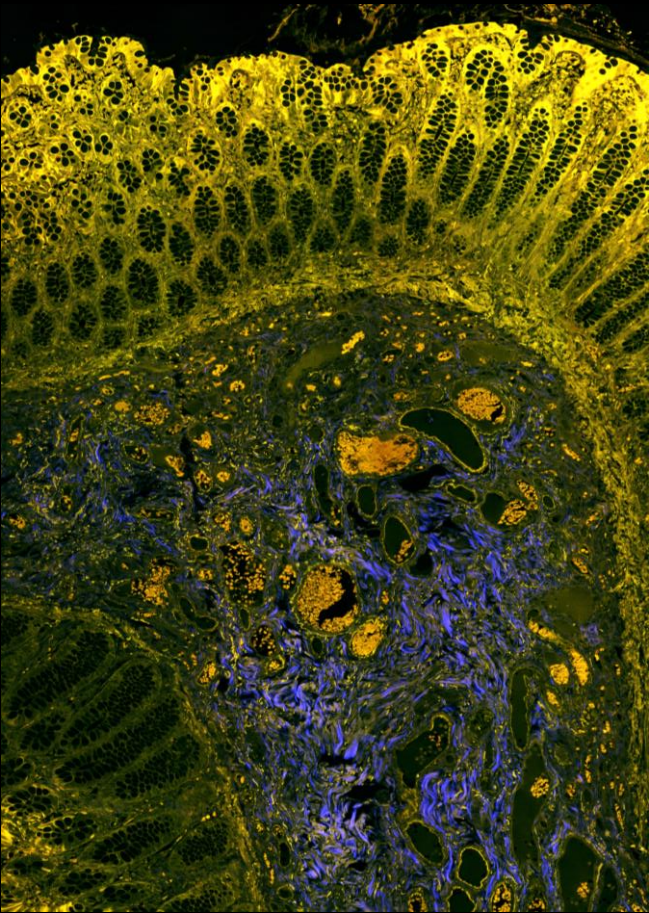
# EXAMPLE: WIDEFIELD GUIDED MULTIPHOTON IMAGING



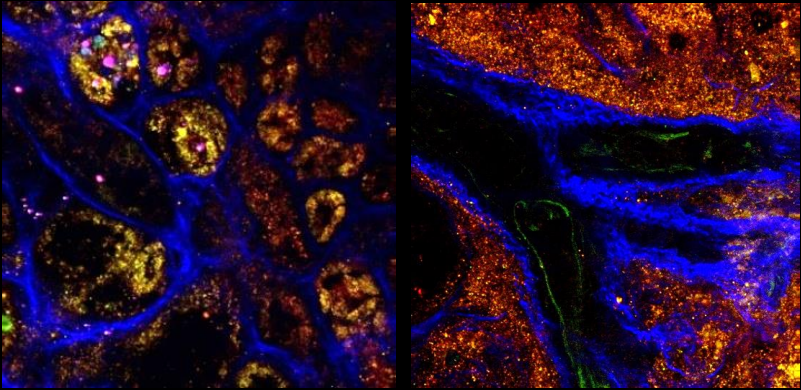


# EXAMPLE: BIOMARKERS AND LABEL-FREE IMAGING: COLLAGEN

## Collagen in 2D Tissue Sections

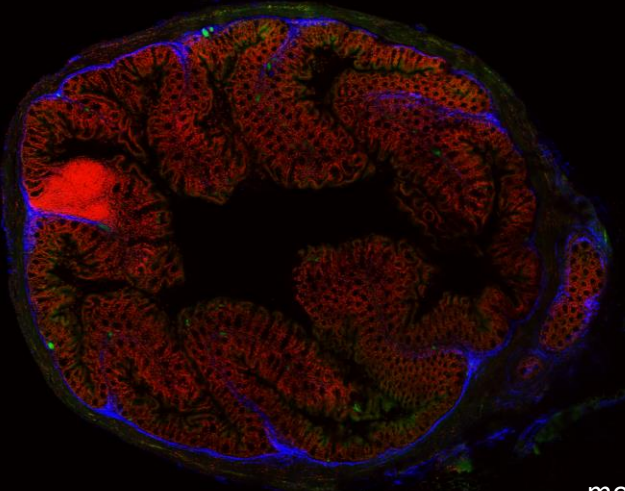
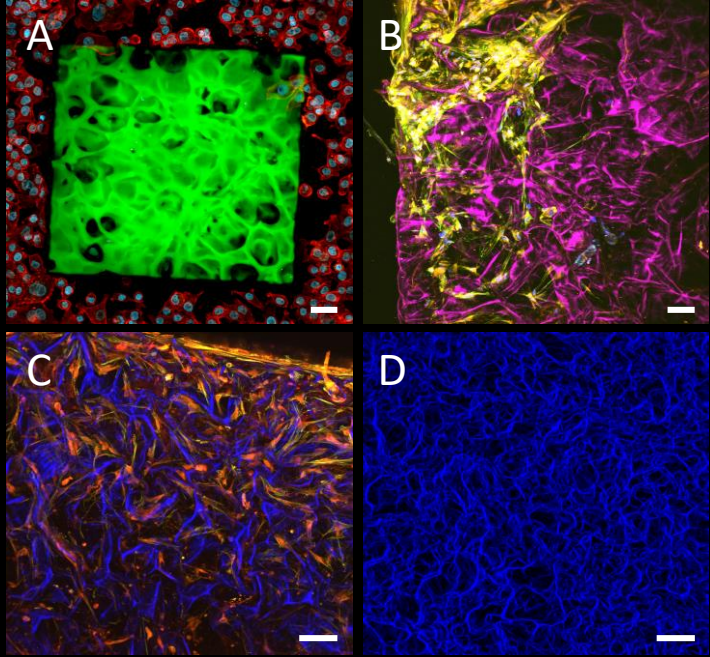


## Collagen in 3D Tissue



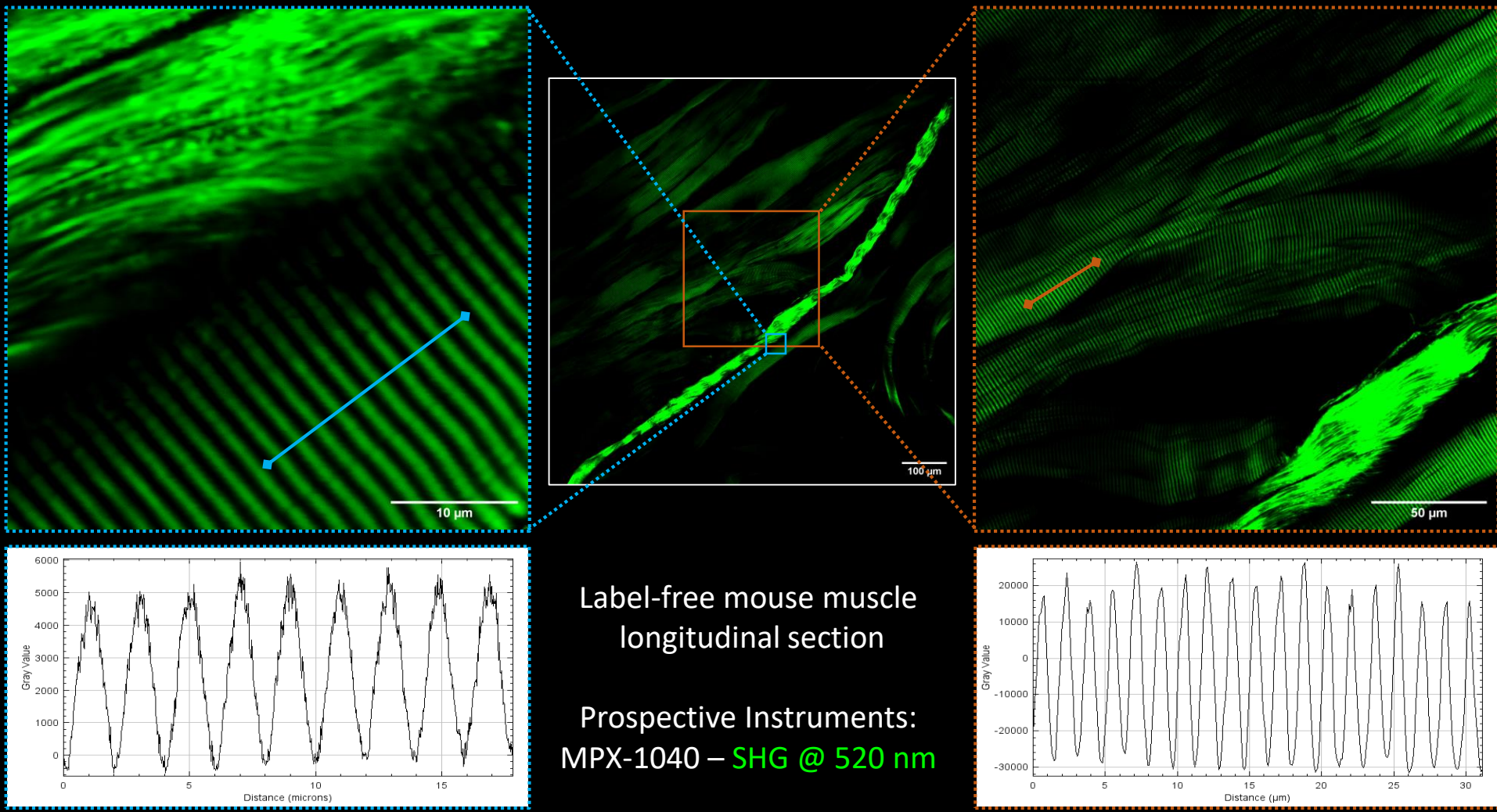
swine kidney

## Collagen-Scaffolds



mouse colon

# EXAMPLE: BIOMARKERS AND LABEL-FREE IMAGING - MUSCLE

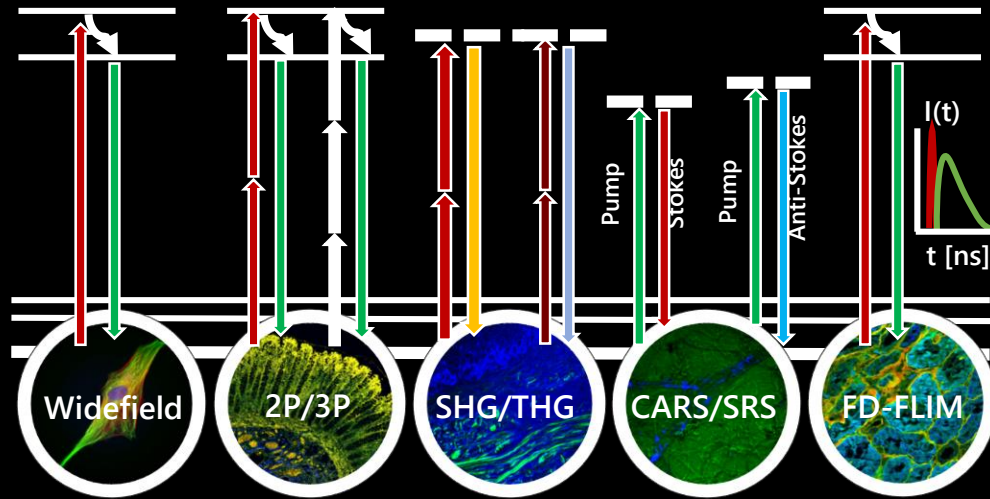


Label-free mouse muscle longitudinal section  
Prospective Instruments: MPX-1040 – SHG @ 520 nm



**TURN-KEY  
EASY-TO-USE  
COMPACT  
MULTIMODAL**

# MULTIMODAL MICROSCOPY



- ✓ Intrinsic confocal
- ✓ Allows optical sectioning → 3D volume scans
- ✓ NIR fs-Lasers allows higher penetration depth
- ✓ NIR fs-Lasers: less photodamage
- ✓ Multiplexing up to 4 channels simultaneously
- ✓ Multimodality: orthogonal data



**TWO-PHOTON, SHG & THG  
OPTOGENETICS  
FLIM  
CARS & SRS**

Multiphoton Microscopes

Femtosecond Lasers

# REFERENCES

## Next-Generation Laser Scanning Multiphoton Microscopes are Turnkey, Portable, and Industry-Ready

Michael E. Holmes,<sup>1\*</sup> Stefanie Kiderlen,<sup>2</sup> and Lukas Krainer<sup>2</sup>

<sup>1</sup>Two-level systems, 501 1st Ave. North, # 901, St. Petersburg, FL 33701  
<sup>2</sup>Prospective Instruments LK OG, Stadlstraße 33, 6850 Dornbirn, Austria  
 \*mholmes@two-levelsystems.com

2 Microscopy TODAY doi:10.1017/S155192522000657 www.microscopy-today.com • 2022 May



## Selective CNS Targeting and Distribution with a Refined Region-Specific Intranasal Delivery Technique via the Olfactory Mucosa

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**TUM** TWO-PHOTON MICROSCOPY AND ACOUSTIC TRAPPING FOR THE ANALYSIS OF CELL OSMOSIS  
 Bettina Sailer<sup>1</sup>, Thomas Kollner<sup>1</sup>, Rune Bärnköb<sup>1</sup>, Thomas Hellner<sup>1</sup> and Oliver Hayden<sup>1</sup>  
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**Abstract**  
 Acoustic trapping of red blood cells (RBCs) and lung cells (LNCs) in a spherical microtrapper (MT) are demonstrated for the investigation and analysis of cellular microviscosity under microscopy methods (lightical and two-photon laser fluorescence (TPLF)). We describe a platform technology (PT) for non-invasive monitoring of cells in 3D without a cell contact in microfluidic flow conditions and compare both microscopy methods for spatial analysis of cell function and cell interactions using osmosis in RBCs. As example, a focus shift to a cell surface combined with TPLF will be discussed to study cell function diagnosis covering a diagnostic gap of in-vitro cell testing in 3D for biomedical research.

**Acoustic Particle Trapping and Cell Osmosis**  
 Comparison of bright field and TPLF microscopy method in acoustic, particle-trapping using osmosis

**Chip Design and Platform Technology**  
 We applied the 3-D, acoustic to perform platform technology in acoustic particle-trapping in 3D.

**Acoustofluidics platform technology**

**Outlook**  
 Our further research will be the expansion in analysis of different cell types towards cell function diagnosis in combination with TPLF for field of biomedical research covering a diagnostic gap of in-vitro cell testing in 3D.

**Conclusion**  
 In this work we have introduced acoustic cell trapping with the example of osmosis in RBCs and lung cells (LNCs) in a spherical microtrapper under two microscopy methods for potential biomedical research applications – lightical and TPLF.

**References**  
 [1] B. Sailer, et al., Microfluid. 2020, 04-08, October 2020, Virtual Conference, Abstract No. 2029  
 [2] D. A. Russell, American Journal of Physics, 76, 549-554 (2008).

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A multiphoton 3D stack image of a whole *Drosophila* eye fly brain.

### A Multiphoton Microscope Enables Portable 3D Biological Imaging

The mobile design featuring built-in laser scanning provides key insights into animal tissue samples and offers a window into human systems to help guide treatment.

BY MICHAEL E. HOLMES AND MARCO FOMINI, NICOLAUS EITZINGER, AND STEFANIE KIDERLEN AND LUKAS KRAINER, PROSPECTIVE INSTRUMENTS

For decades, optical microscopy has provided the mechanism with which to image cells and tissues for the purposes of basic research, digital pathology, and the study of the brain. Samples are typically studied in tissue sections. Most of these systems use confocal microscopy and single-photon excitation, typically from a continuous-wave light source, to excite the sample. Multiphoton microscopy is used when deep-tissue and cellular-level resolution are desired to preserve the integrity of the tissue to image cells and tissues for scanning multiphoton microscopes. Light from an external laser is tightly focused and scanned across the sample using fast mirrors. An image is created by detecting the fluorescence signal intensity at each point and mapping spatially with the aid of computer software. The excitation source and signal detection implementation are critical for biological imaging, ease of use, and the quality of the microscope. Multiphoton microscopy uses a tightly focused laser beam in the near-infrared (NIR) range, typically between 700 and 1300 nm, to produce nonlinear optical effects based on the interaction of multiple photons exciting chromophores in a molecule. Therefore, the intensity of the generated signal does not increase linearly with the excitation source and signal.

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Biopixlar® Application Note

## Microfluidic Spheroid Bioprinting for Human Disease Modeling

Dr. Andreas Svanström,  
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**Figure 3. A)** HepG2 cells cultured for 4 days post printing and gel embedding, imaged using phase microscopy. **B)** HepG2 and 3T3-J2 cells cultured for 7 days post printing and gel embedding, imaged using 2-photon microscopy. Spheroids cytoskeleton and nuclei stained using Actin-Phalloidin (AF-488, green) and propidium iodide (red), respectively. Image shows maximum intensity projection. (https://www.p-inst.com/)

Artikel | Mikroskopie Lichtmikroskopie 3 April 2023

# Turnkey Multiphoton Microscopy for 3D samples

Label-free and 4D imaging in Life Science and Medicine

Stefanie Kiderlen, Lukas Krainer

Wiley Analytical Science Award Winner

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