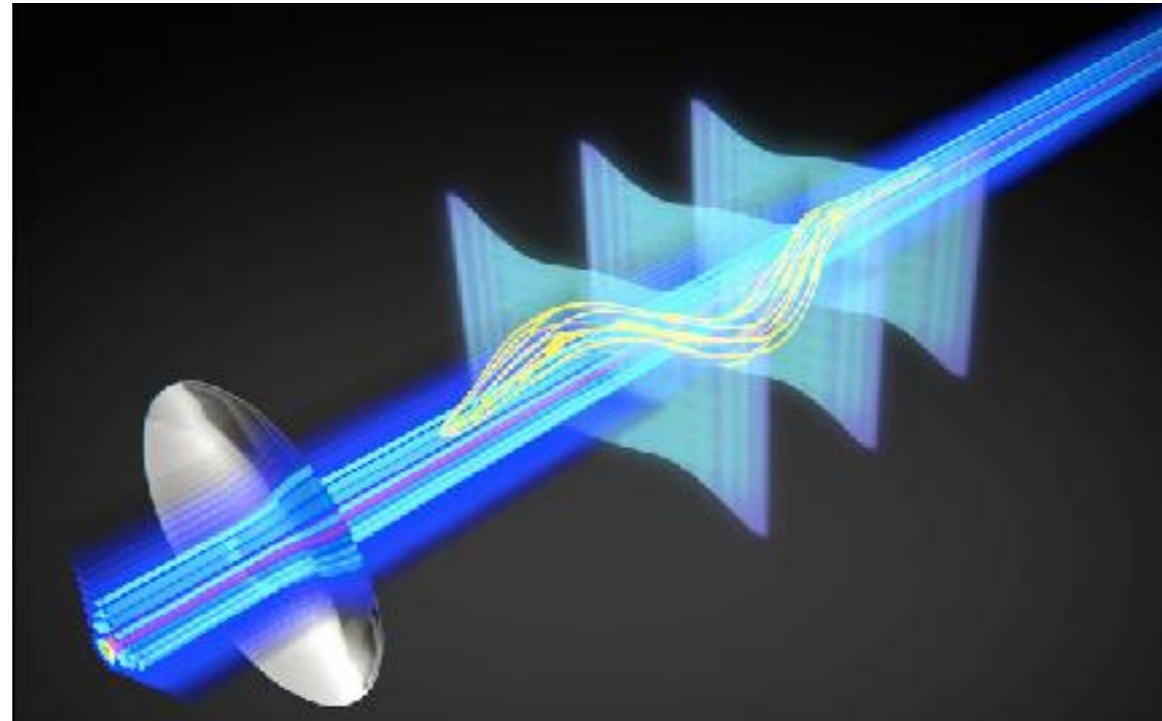


Challenges in Two-photon Excited Microscopy and in Light Sheet Microscopy



The Super resolution light microscopy
and nanoscopy lab (SLN)

Pablo Loza-Alvarez

pablo.loza@icfo.eu

<http://sln.icfo.eu>

The Super resolution light microscopy and nanoscopy lab (SLN)

ACCESS

User driven

(biological applications)

**Improved Specs
Unique Features
Custom made**

R&D

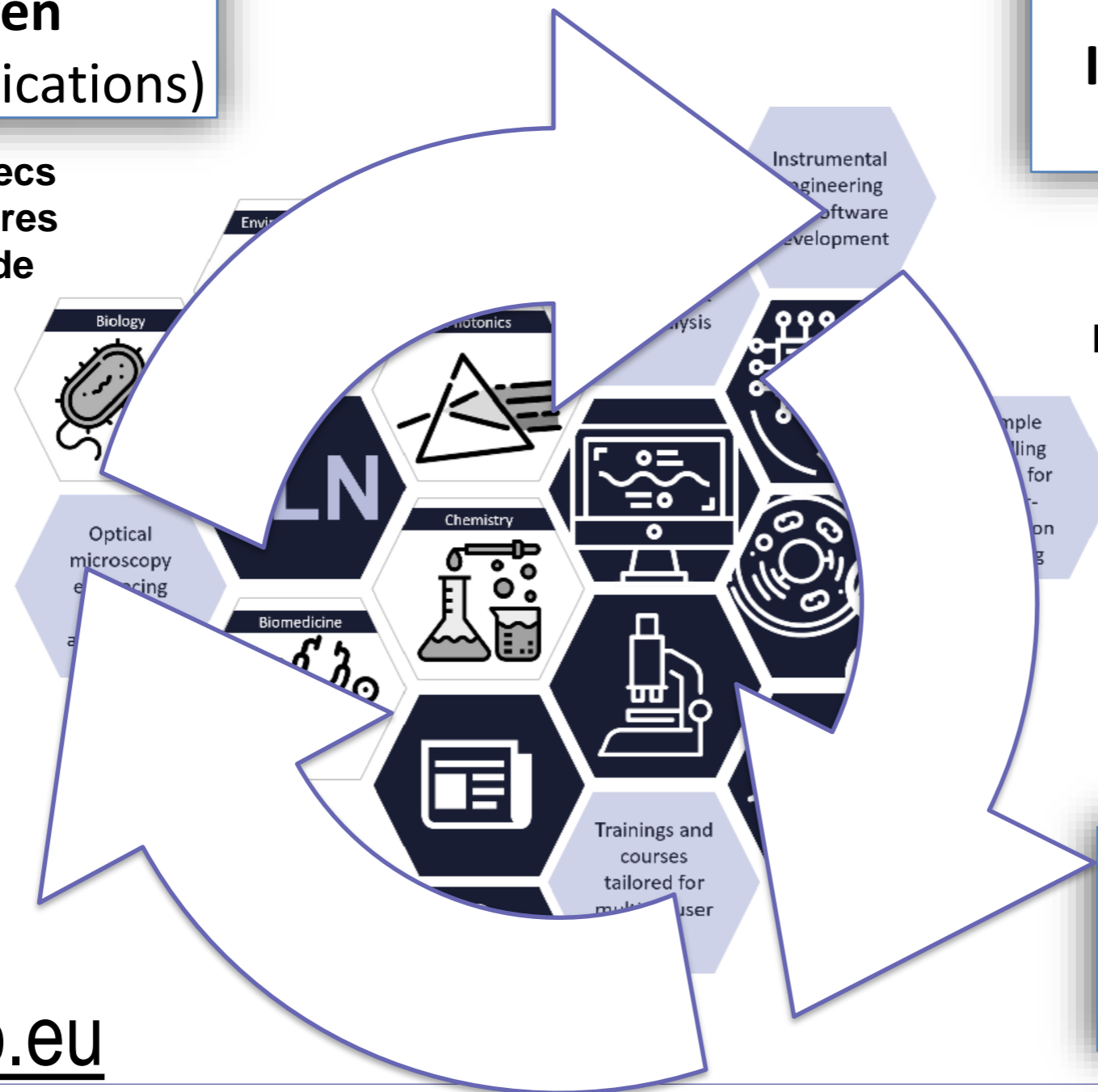
**Instrument development
(optics and photonics)**

**Adapt: new technologies
Merge: imaging techniques
Develop: novel instrumentation**

**New Generations
Different Backgrounds
According to Needs**


TRAINING

**Latest technologies
(all types of users)**




<http://sln.icfo.eu>


The SLN Team



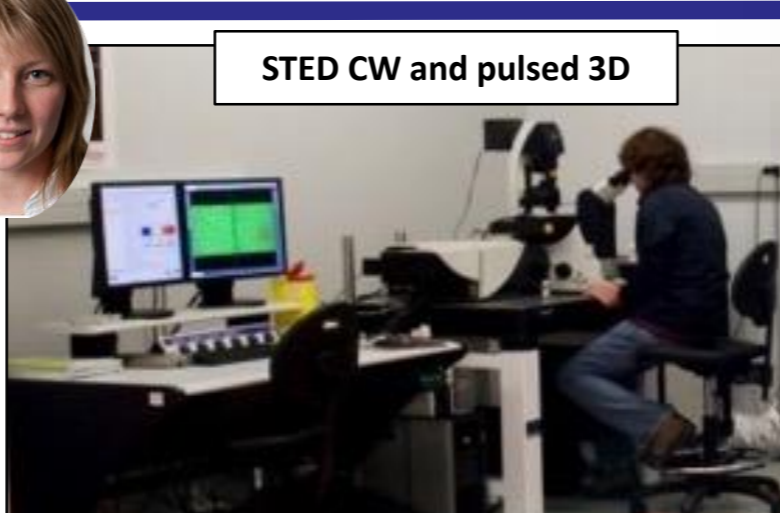
STORM




**EMCCD (high sensitivity) + sCMOS (fast imaging)
+ variable FOV + NIS v5 software**



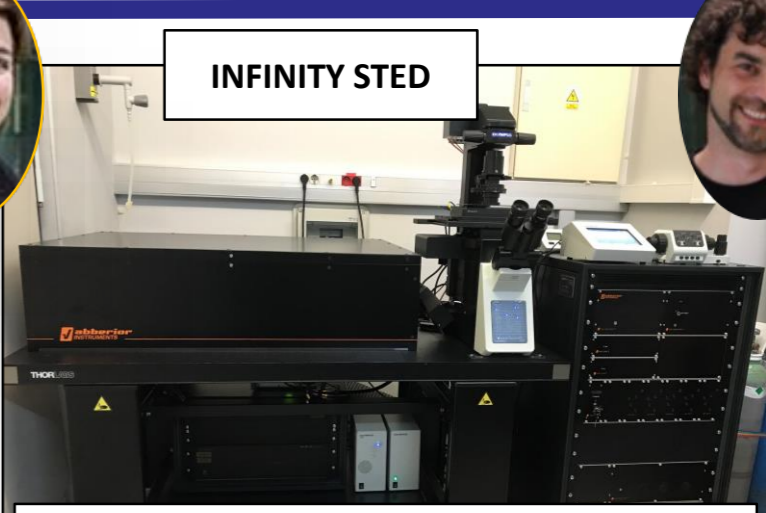
STED CW and pulsed 3D






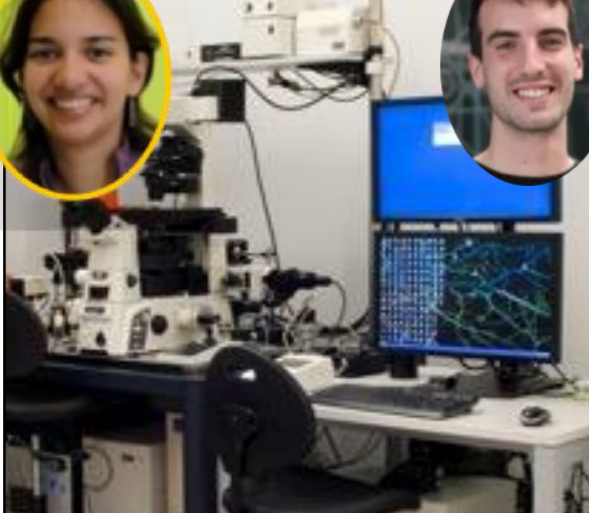
**FLIM (tauSTED) + Multimodal
+ novel laser sources + novel algorithms**



INFINITY STED



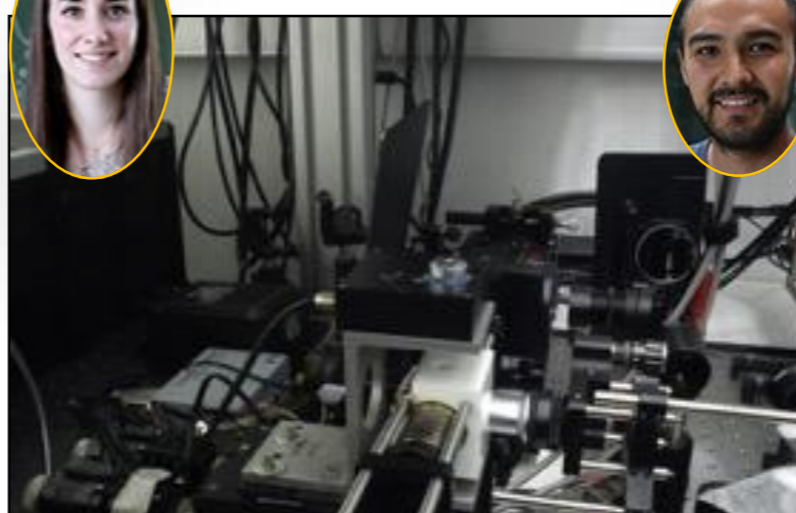


DyeMIN + Rescue + Minfield + easy 3D + FLIM

Confocal and multiphoton

Spectral Confocal+
TPEF +SHG+THG (nonlinear)
+AOSLO+ Photomanipulation

4 Light Sheet Microscopes

Linear, nonlinear, RAMAN
High throughput, Fast 3D imaging





RAMAN

Advanced Mathematical and
Statistical analysis

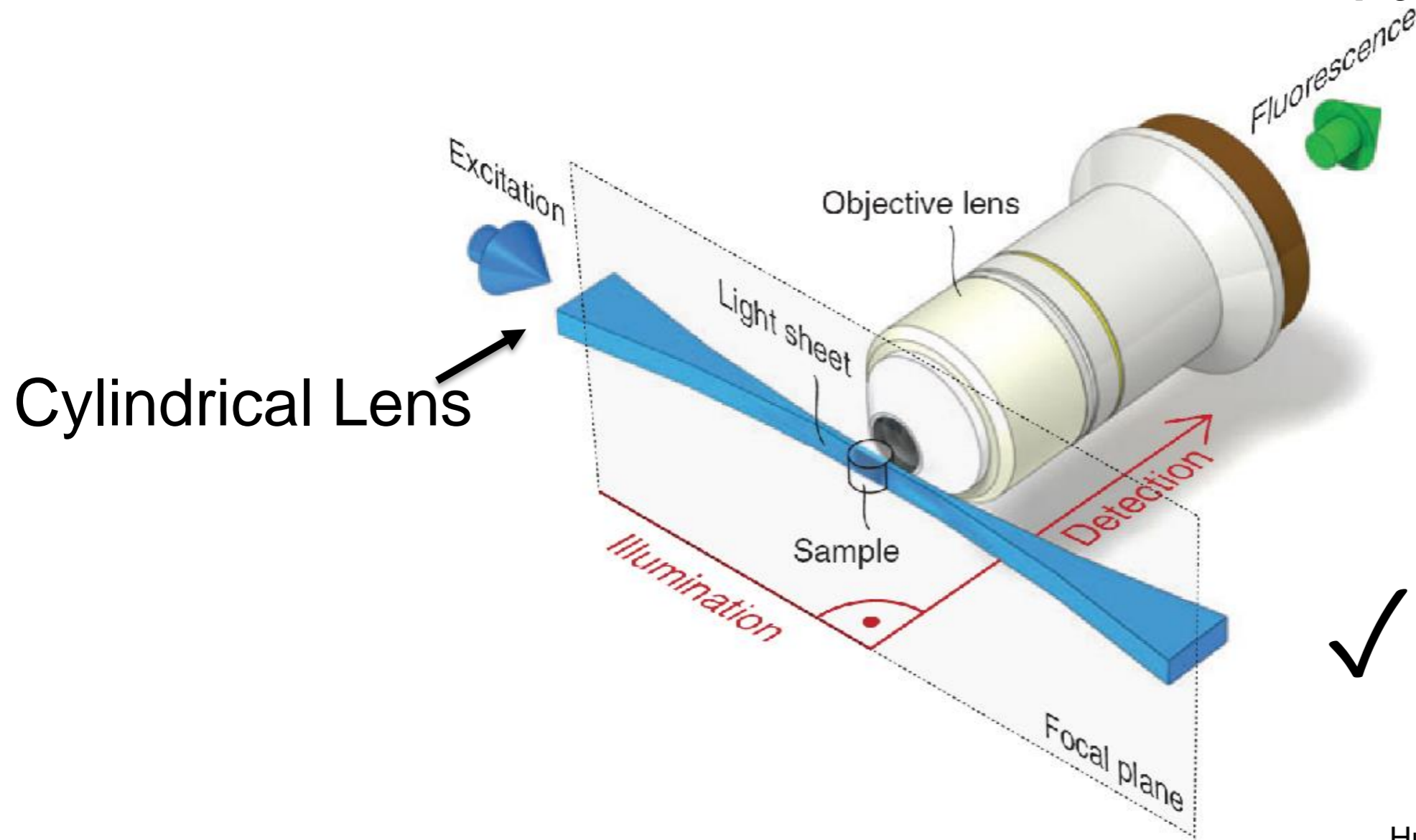
- 10 researchers:
- 3 Permanent Staff
- 4 Postdocs
- 3 PhD students

- EU, National and Regional projects
- Private and public funded
- ~10+ Q1 pub/year

- 11 imaging systems
- <80 Internal and External users/year
- Usage/year: 8K-9K hrs

Light-Sheet fluorescence Microscopy

Selective Plane Illumination Microscopy (SPIM)



✓ FAST imaging

Adapted from
Huisken, J. et al., Development (2009)

Light-Sheet fluorescence Microscopy

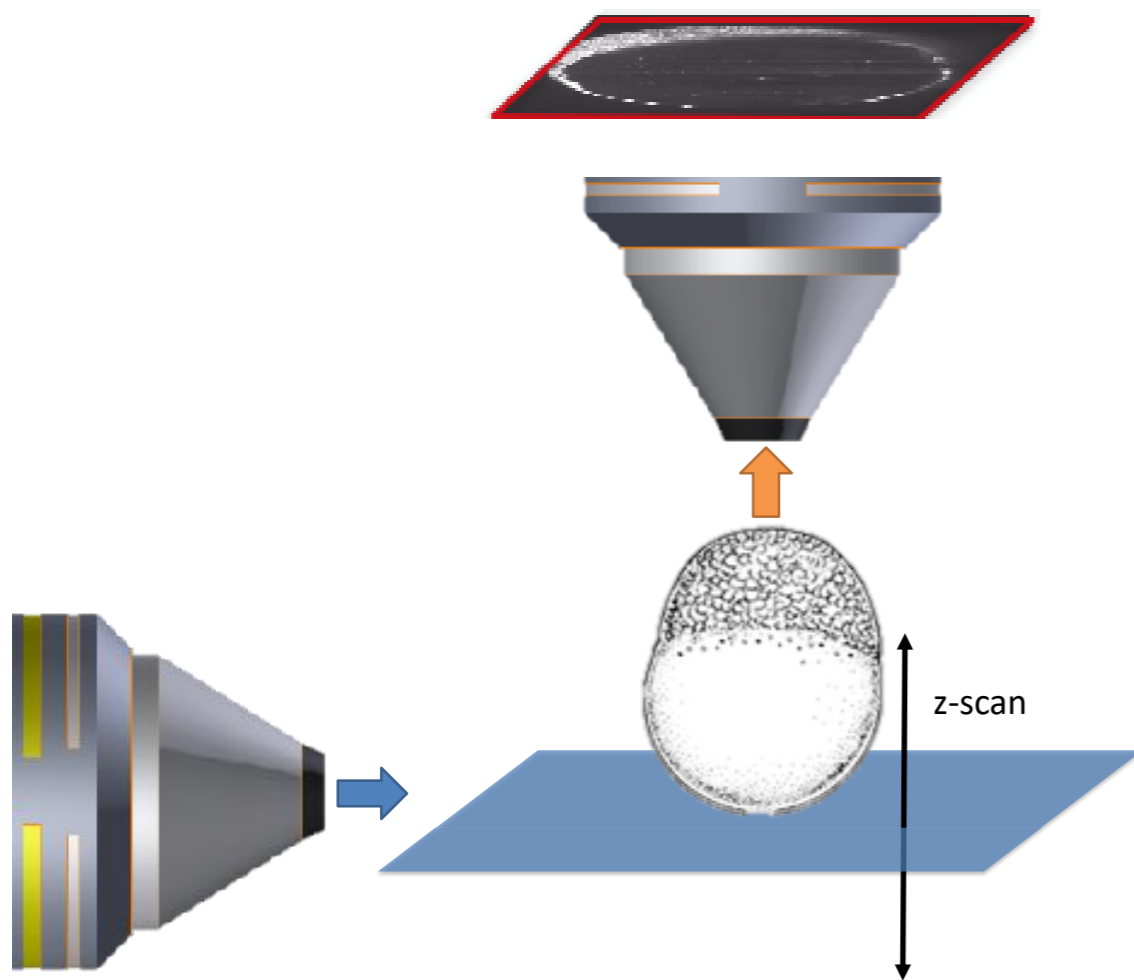
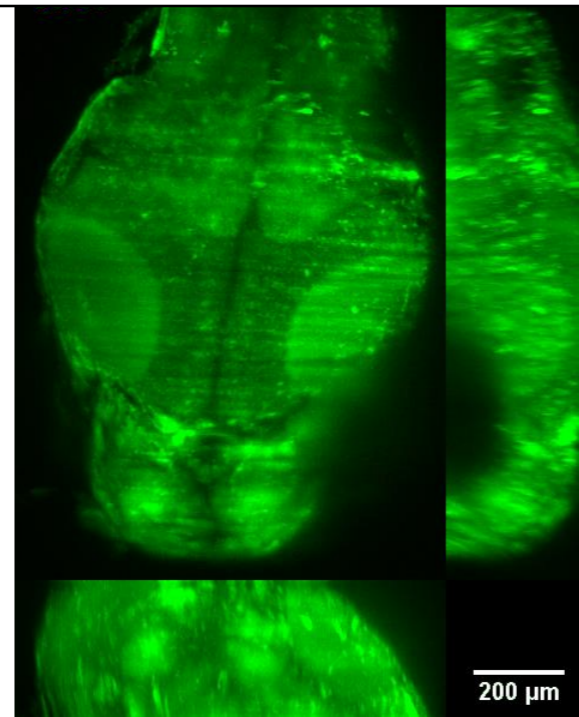
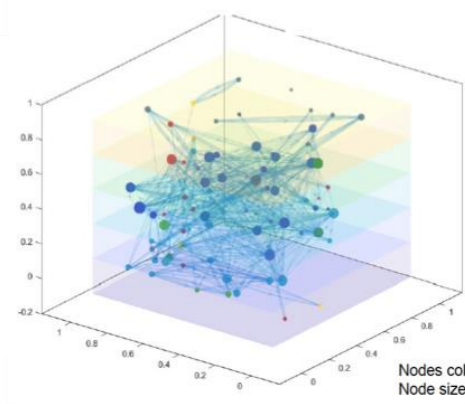
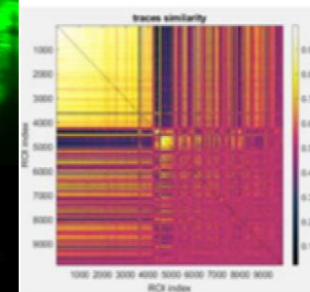


Image speed ~ 1 hertz



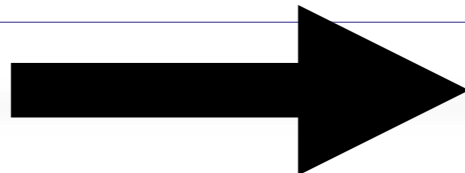
Zebra fish brain activity
Top, lateral and frontal projections

3D Connectivity maps



A 3D image is formed by:

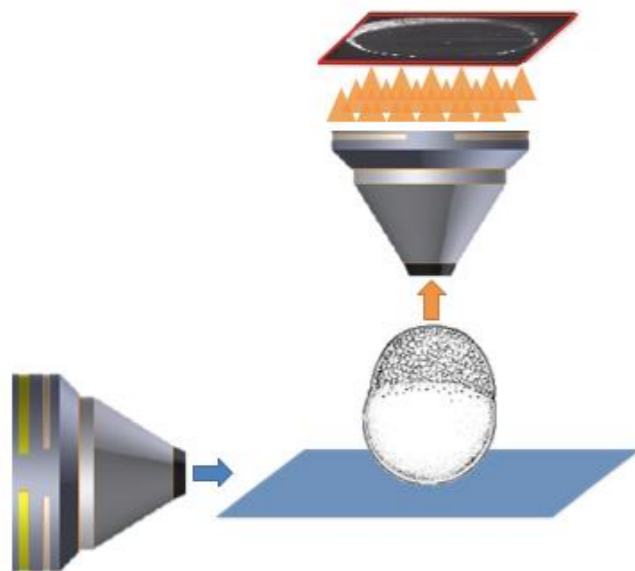
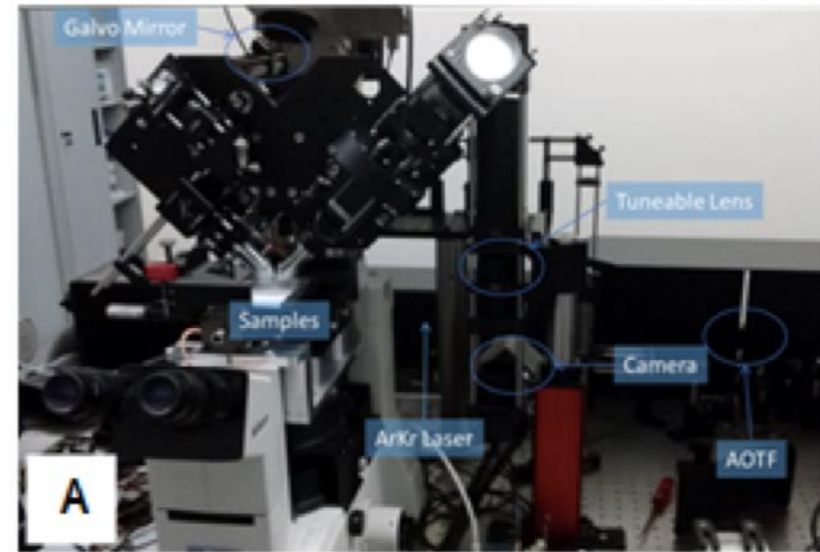
MOVING the **SAMPLE**
through
the light sheet



Important dynamics may be lost!

Fast volumetric imaging in LSFM

I: Use Electrically tunable lenses

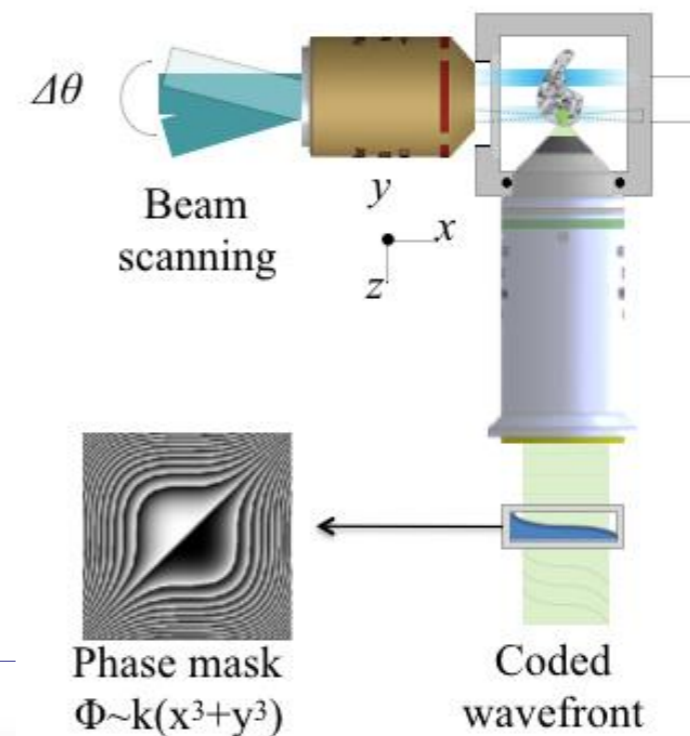


A 3D image is formed by:

MOVING the **SAMPLE**
through
the light sheet

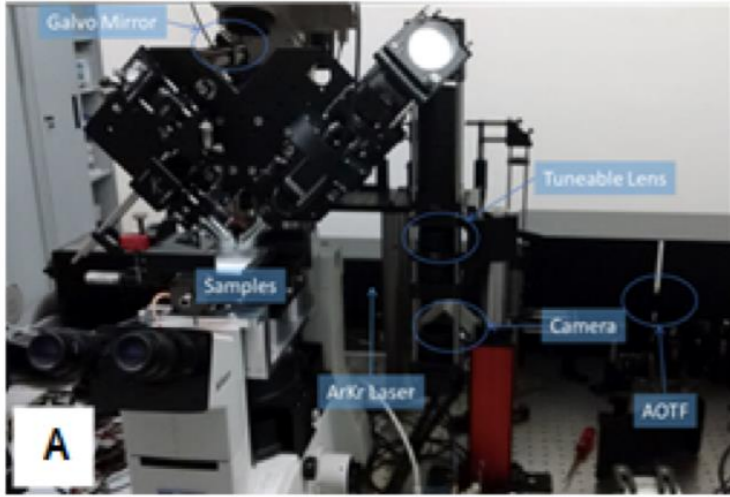
SOLUTION:
ONLY Move the light sheet

II: Extending Depth of field (wave front coding)



Fast volumetric imaging in LSFM

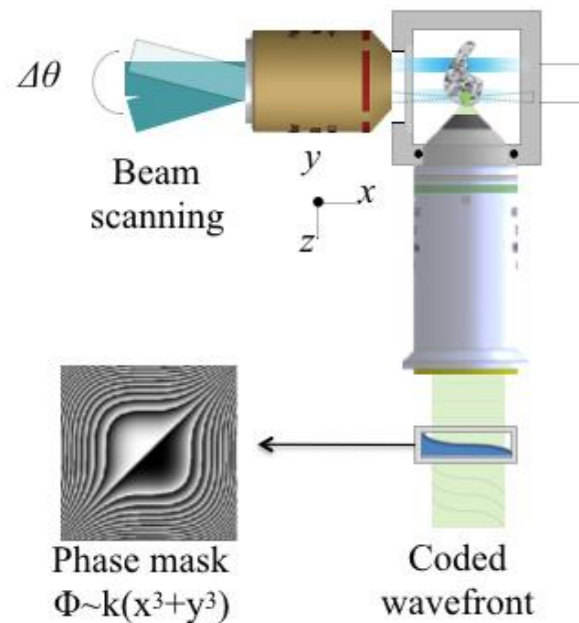
I: Use Electrically tunable lenses



25 vols/sec
Small volumes
1Vol < 10 planes

At fast speeds
Loss of linearity
Loss of amplitude

II: Extending Depth of field (wave front coding)



Potential to reach 100 V/s

Needs calibration (PSF)
Requires deconvolution
(high SNR)

CHALLENGES

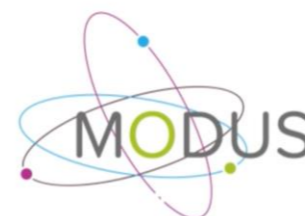
- Brighter Fluorescent markers
- Faster 2D array detectors
High resolution (small pixels)
High sensitivity
- Large amount of data
(~0.5 TB/hr)
Storage
Processing
Accessibility



PHOTONICS²¹



ADVANCED MULTIMODAL PHOTONICS LASER IMAGING TOOL FOR UROTHELIAL DIAGNOSIS AND ENDOSCOPY (AMPLITUDE)

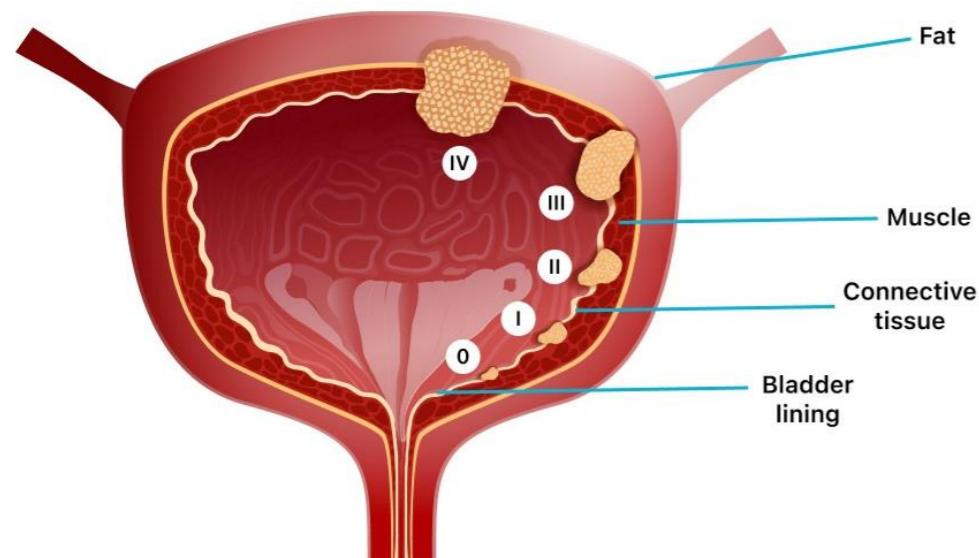


www.amplitude-imaging.com

Bladder Cancer

To identify the first stages (0, I) of bladder cancer

Stages of Bladder Cancer



5-year survival rate decreases with increasing stage:

0, I → 82-100%;

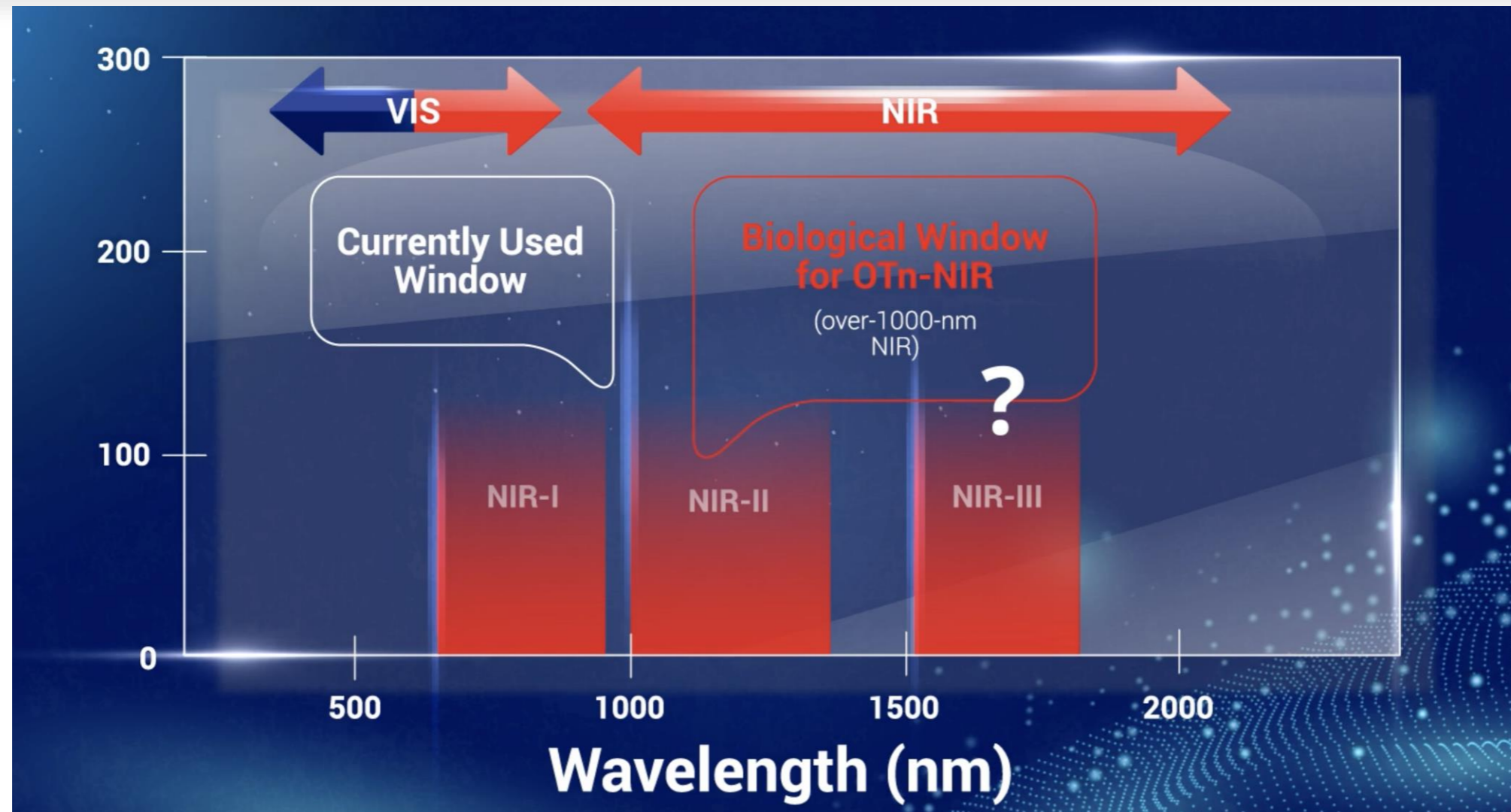
II → 63-83%;

III → 17-71%;

IV → 0-22% (*despite radical treatment*)

Bladder thickness ~ 3mm,

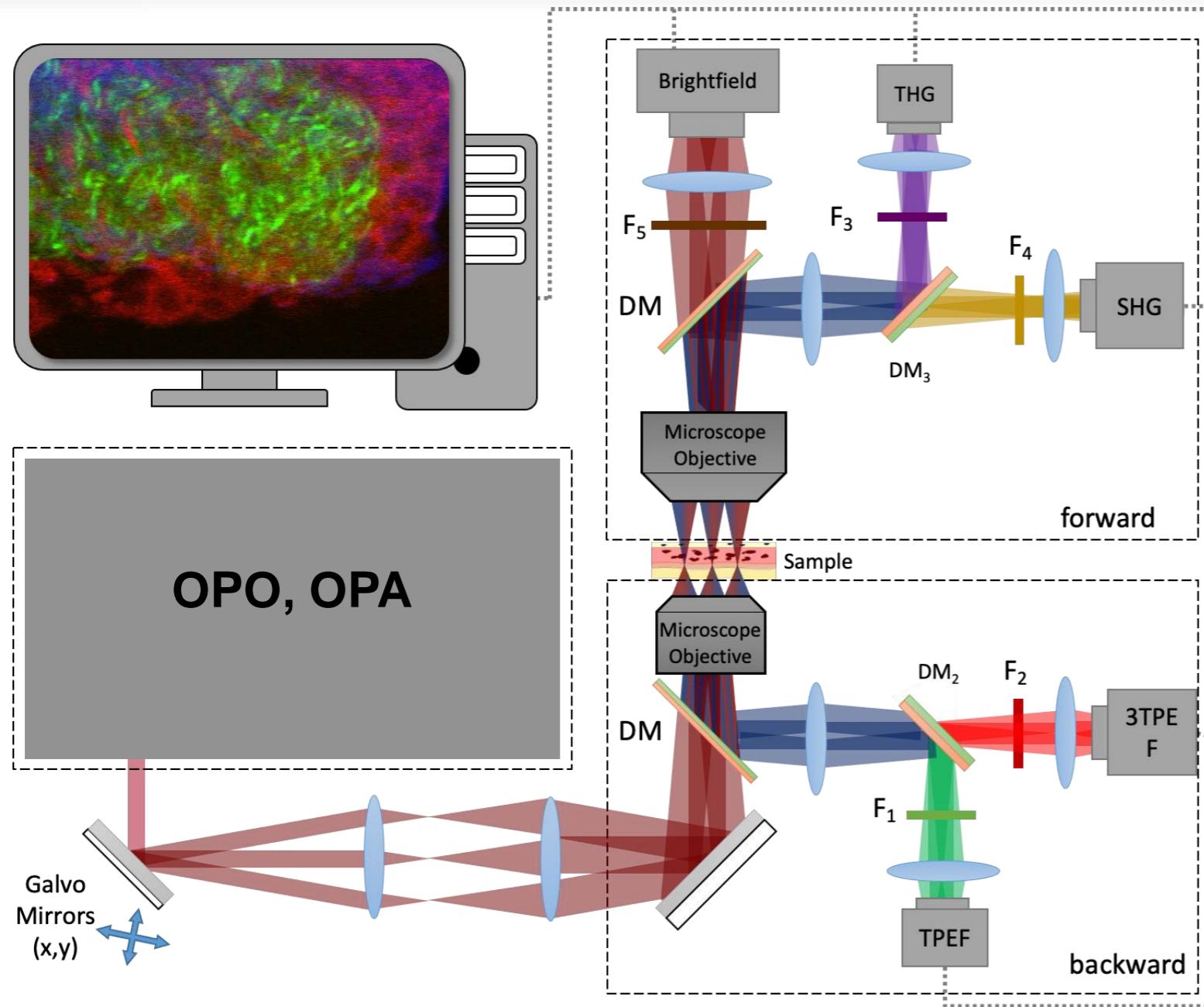
Explore the third optical window for imaging at large penetration depths



Stretched ~1mm

Optically accessible at 1700nm

Label free imaging: NON LINEAR



Explore the use of the OPO at 1700nm for

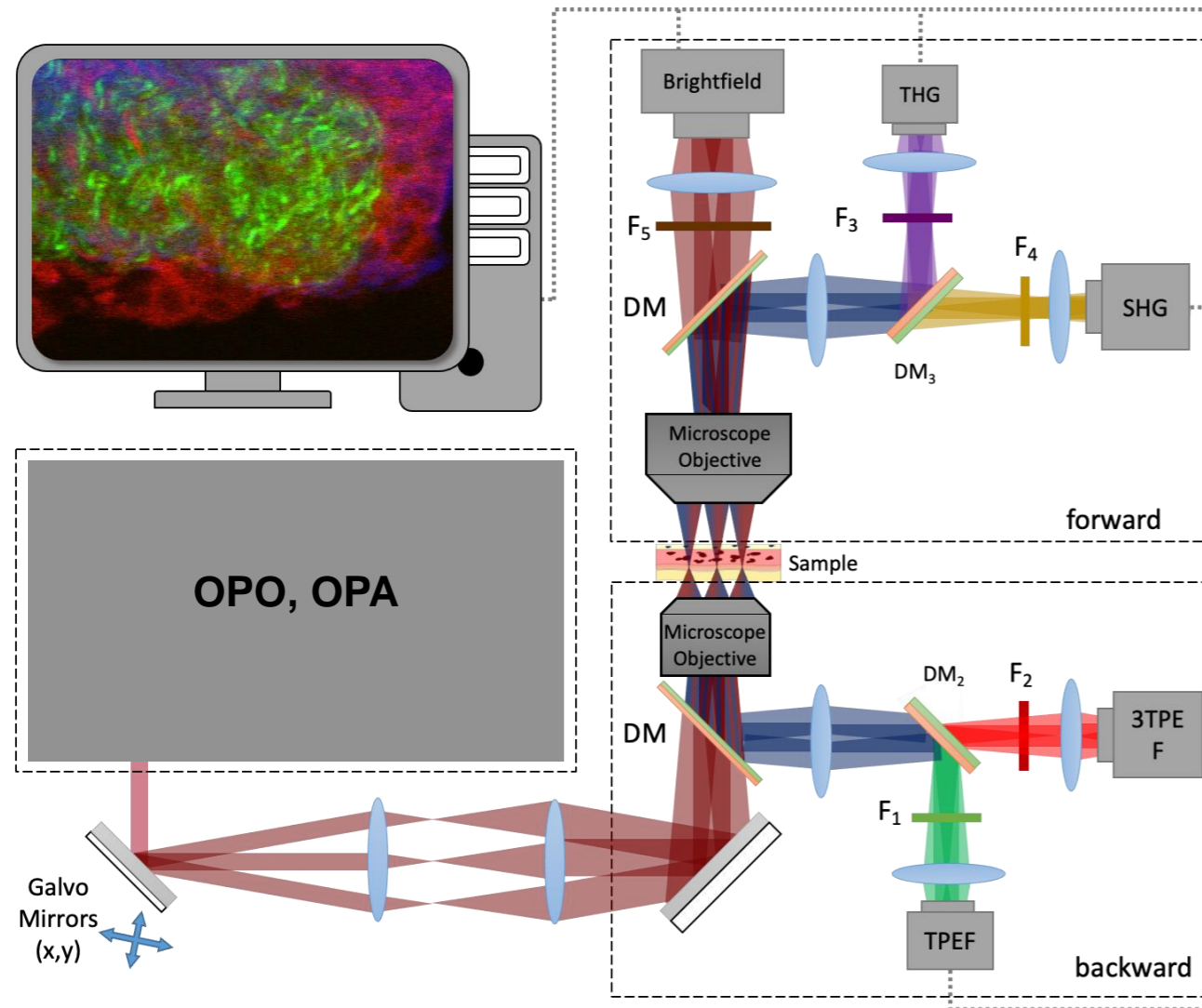
- THG (566nm),
- 3PEF (Visible autofluo),
- TPEF (autofluorescence),
- SHG (850nm),
- Elastic scattering

Frequency doubled OPO source (at 850 nm)

- TPEF (autofluorescence),
- SHG (425nm),
- elastic scattering

Challenges and opportunities

CHALLENGES



OPPORTUNITIES

Thank you!

